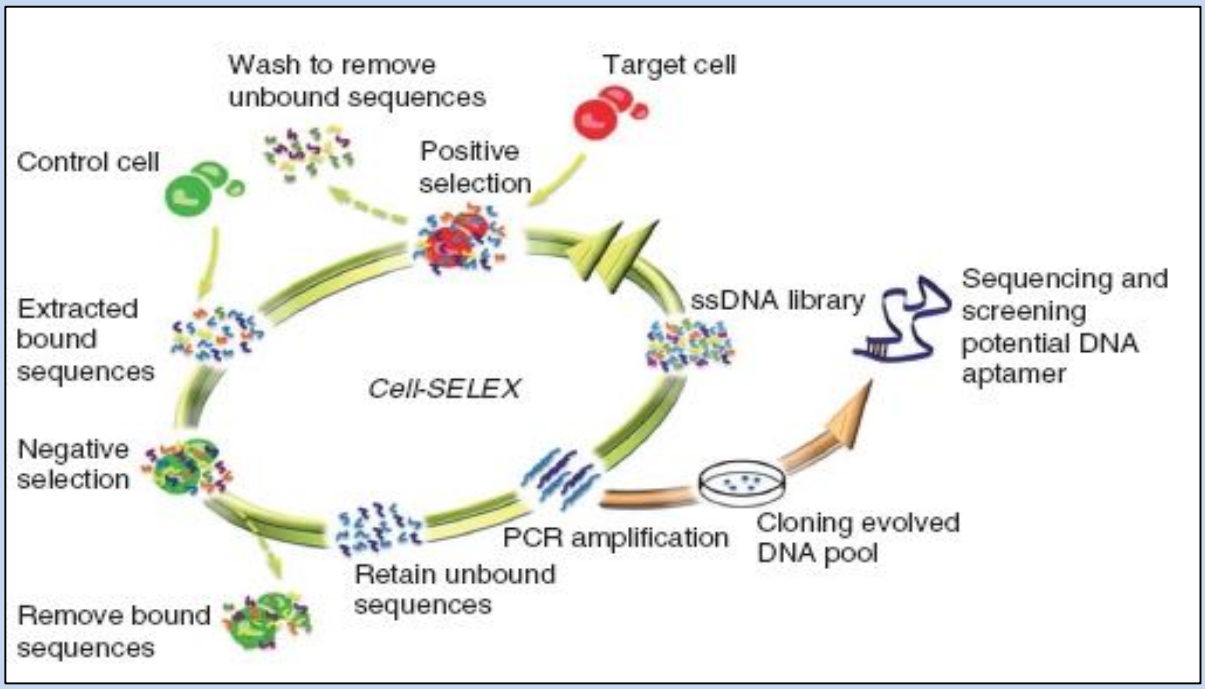
Protocol for Cell-SELEX



1. ssDNA pools are dissolved in 200 μl of PBS binding buffer, denatured at 95 °C for 5 min, and cooled immediately on ice for 10 min before selection.
2. The denatured ssDNA pools is then incubated with 1-2×10^4 target cells in a humid atmosphere at 37 °C for 1h.
3. After washing, the bound DNAs are eluted by heating at 95 °C for 5 min in 200μl of binding buffer.
4. The eluted DNAs were then incubated with negative cells (5-fold excess than target cells) at 37 °C for counter selection for 1 h.
5. After centrifugation, the supernatant is desalted and amplified by PCR.
6. PCR products of unequal length are separated and purified by electrophoresis in a 10% polyacrylamide-7M urea gel to get sense ssDNA.
7. Detect the combination rate of ssDNAs with indicating cells using flow cytometry.
8. Repeat the above steps until no significant further changes in fluorescence intensity are observed.